

Evaluation of the Effect of MEDG Stem Bark on Oxidative Stress in Rat Plasma Caused by Diabetes Mellitus

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ABSTRACT

Diabetes mellitus (DM) is a serious health condition which reduces the quality of life of sufferers. The different treatment options for DM cannot effectively ameliorate the various complications caused by the disease. The aim of the present study was to evaluate the effect of methanol fraction of ethanol extract of *Dialium guineense* (MEDG) stem bark on oxidative stress in rat plasma caused by DM. Male albino rats (Wistar strain; n = 25; mean weight = 215 ± 15 g) were divided into five groups (5 rats per group): control, diabetic, metformin, and 200 mg/kg body weight (bwt) and 300 mg/kg bwt MEDG groups. Diabetes mellitus was induced in the rats via intraperitoneal injection of streptozotocin (STZ, 50 mg/kg bwt). The diabetic rats were subsequently treated with metformin (50 mg/kg bwt) or the medicinal plant extract (200 and 300 mg/kg bwt, respectively), for 21 days. Markers of oxidative stress were measured in rat plasma. The results indicated that the blood glucose concentrations of the rats elevated by STZ-induced DM was significantly reduced after treatment with MEDG stem bark ($p < 0.05$). Similarly, activities of all the antioxidant enzymes [catalase, superoxide dismutase (SOD), glutathione peroxidase (GPx), and glutathione reductase (GR)], and concentrations of glutathione (GSH) were significantly lower in diabetic group than in the control group, but they were increased by MEDG treatment ($p < 0.05$). However, the concentrations of nitric oxide (NO) and malondialdehyde (MDA) elevated by STZ were greatly reduced after treatment with the medicinal plant extract ($p < 0.05$). The results obtained in this study have shown that MEDG stem bark can stimulate antioxidant defense system in rats challenged with STZ.

Keywords: *Dialium Guineense*; Free Radicals; Glutathione Peroxidase; Nitric Oxide; Oxidative Stress

Abbreviations: ROS: Reactive Oxygen Species; RNS: Reactive Nitrogen Species; PKC: Protein Kinase C; DM: Diabetes Mellitus; GR: Glutathione Reductase

Introduction

Implicated in the pathogenesis of many diseases, oxidative stress contributes significantly to insulin resistance [1,2]. Impairment of insulin signaling pathway by reactive oxygen species (ROS) and reactive nitrogen species (RNS) have been reported by different studies [3]. Oxidative stress plays a crucial role in the development of diabetic complications (micro- and macrovascular). The metabolic abnormalities of the disease cause mitochondrial overproduction of superoxide anion in endothelial cells of both large and small vessels, and also in the myocardium. This in turn causes the activation of five major pathways involved in the pathogenesis of diabetic complications: polyol pathway flux, increased formation of advanced glycation end-pro-

ducts (AGEs), increased expression of the receptor for AGEs and its activating ligands, activation of protein kinase C (PKC) isoforms, and overactivity of the hexosamine pathway. It also directly inactivates two important antiatherosclerotic enzymes, endothelial nitric oxide synthase (eNOS) and prostacyclin synthase. Through these pathways, increased intracellular ROS cause defective angiogenesis in response to ischemia, activates a number of pro-inflammatory pathways, and cause long-lasting epigenetic changes which drive persistent expression of pro-inflammatory genes after glycemia is normalized (hyperglycemia memory) [4-6]. In pancreatic β -cells, NO regulation of glucokinase activity via S-nitrosylation reaction, may enhance insulin secretion [7,8].

However, excess NO and concomitant NRS could cause apoptosis through caspase-3 activation and decrease in ATP levels [9]. Studies have speculated a direct relationship between the health benefits of a plant and its antioxidant content. *Dialium guineense* (Velvet Tamarind) is a medicinal plant used in Traditional Medicine for the treatment of infections [10]. It is a tall, tropical, fruit-bearing tree, belonging to the Leguminosae family, and has small, typically grape-sized edible fruits with brown hard inedible shells. In Africa, it is found in dense forests along the southern edge of the Sahel. The plant grows naturally in West African countries, Central African Republic, and Sudan [11]. In Nigeria, it is known by different local names: Icheku (Igbo), Awin (Yoruba), Tsamiyarkurm (Hausa) and Amughen (Bini) [12]. The aim of this study was to evaluate the effect of MEDG stem bark on oxidative stress in rat plasma caused by diabetes mellitus (DM).

Materials and Methods

Chemicals

Analytical grade reagents were used in this study. Kits used to carry out antioxidant assays were bought from Randox Laboratories Limited (United Kingdom). The other chemicals/reagents were products of Pyrex Scientific Limited (United Kingdom), Merck (Germany), British Drug House (BDH) (England), and Sigma-Aldrich Ltd. (USA).

Plant Material and Authentication

Freshly harvested stem barks of *D. guineense* were collected from Auchu, Etsako West, Edo State, Nigeria. Their identification and authentication took place at the University of Benin herbarium in the Department of Plant Biology and Biotechnology, Faculty of Life Sciences (No. UBHD330).

Plant Preparation and Extraction

The plant stem barks were washed and shade-dried at room temperature for 30 days, and thereafter ground into powder using an electric blender. A portion (500 g) of pulverized plant material was steeped in 5 L of absolute ethanol. The resultant extract was filtered through muslin cloth and freeze-dried with a lyophilizer [13-15].

Experimental Rats

Male rats (Wistar strain, $n = 25$) with weight ranging from 200 to 230 g (mean weight = 215 ± 15 g) were purchased from the Animal House of the Department of Anatomy, School of Basic Medical Scienc-

es, University of Benin, Benin City, Nigeria. The rats were kept in metal cages under standard laboratory settings. They had unrestricted access to feed (pelletized mash) and potable drinking water. Seven days were allowed to acclimate the rats to laboratory conditions prior to commencement of the study. The investigation followed a standard experimental protocol.

Experimental Design

The rats were randomly assigned to five groups (5 rats in a group): control, diabetic, metformin, and 200 mg/kg bwt and 300 mg/kg bwt extract groups. Diabetes mellitus was induced in the rats via intraperitoneal injection of STZ (50 mg/kg bwt). The diabetic rats were then treated for a period of 21 days with metformin (50 mg/kg bwt) or MEDG (200 and 300 mg/kg bwt, respectively), leaving the diabetic group untreated.

Preparation of Plasma

At the end of the 21-day treatment, the rats were euthanized under mild anesthesia. Blood was collected through cardiac puncture into sterile heparin containers. The blood samples were centrifuged at 2000 rpm for 10 min to obtain clear plasma.

Biochemical Analyses

The activities of catalase, SOD, GPx and GR were determined [16-19]. Concentrations of plasma total protein, MDA and GSH were also measured [20-22]. Nitric oxide (NO) concentration was determined as described in literature [23].

Statistical Analysis

Data are expressed as mean $\pm$ SEM ($n = 5$). Statistical analysis was performed using SPSS version 21. Statistical differences between means were compared using Duncan multiple range test. Values of $p < 0.05$ were considered statistically significant.

Results

Effect of MEDG Stem Bark on Weight and Blood Glucose of Rats

As shown in Table 1, the blood glucose concentrations of the rats elevated by STZ-induced DM was significantly reduced after treatment with MEDG stem bark ($p < 0.05$).

Table 1: Effect of MEDG Stem Bark on Weight and Blood Glucose of Rats.

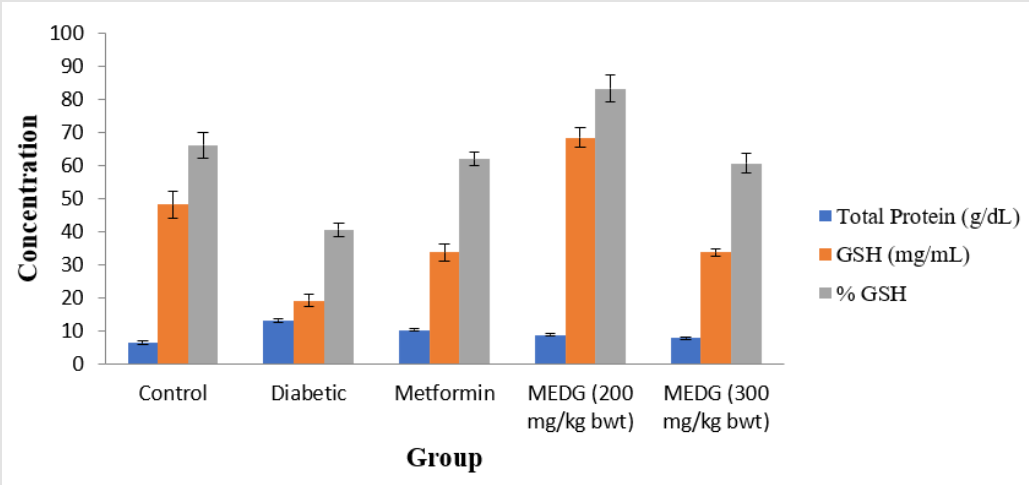
Group	Weight Change (g)	% Weight Change	FBG (mg/dL)	Glycemic Change (mg/dL)	% Glycemic Change
Control	-	-	-	-	-
Diabetic	-	-	> 800	-	-
Metformin	20.35	12.16	> 800	399	49.88
MEDG (200 mg/kg bwt)	12.26	7.87	> 800	421	52.63
MEDG (300 mg/kg bwt)	29.08	17.02	364	227	62.36

Note: Data are weight and FBG parameters and are expressed as mean ± SEM (n = 5).

Oxidative Status of Diabetic Rat Plasma

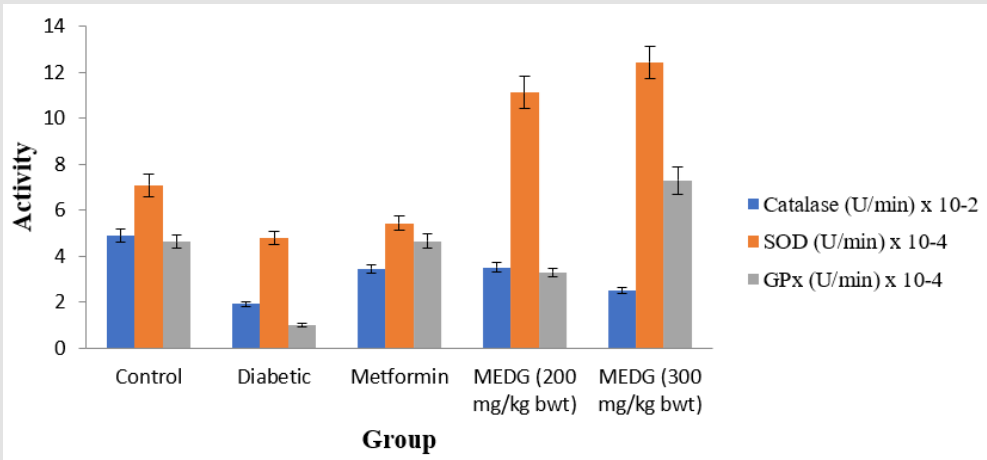
The activities of all the antioxidant enzymes and concentrations of GSH were significantly lower in diabetic group than in the control

group, but they were increased by MEDG treatment ($p < 0.05$). However, the concentrations of NO and MDA elevated by STZ were greatly reduced after treatment with the medicinal plant extract ($p < 0.05$; Figures 1-4).



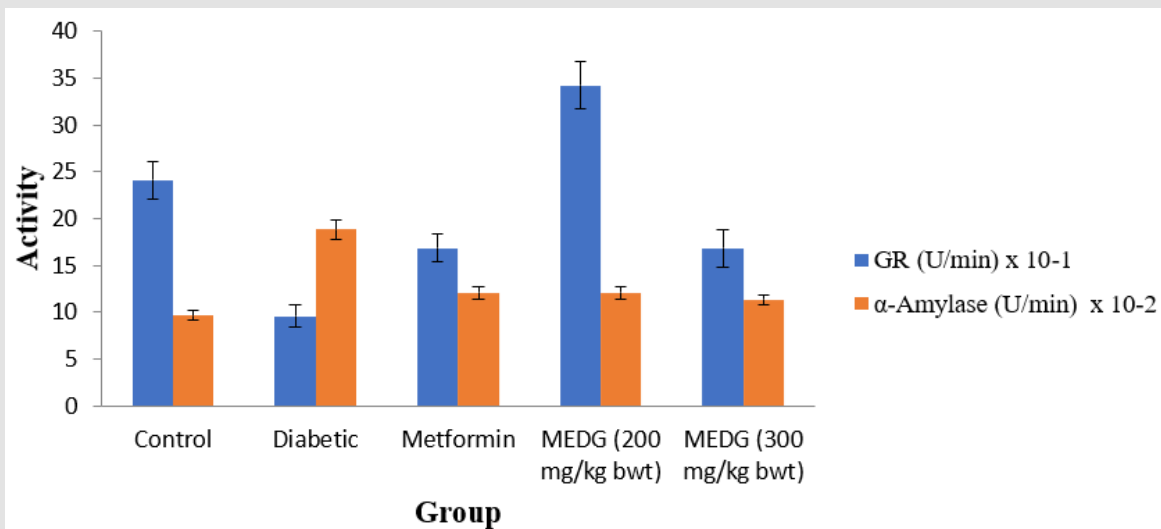
Note: Data are plasma total protein and GSH levels and are expressed as mean ± SEM (n = 5).

Figure 1: Effect of MEDG Stem Bark on Plasma Total Protein and Glutathione Levels.



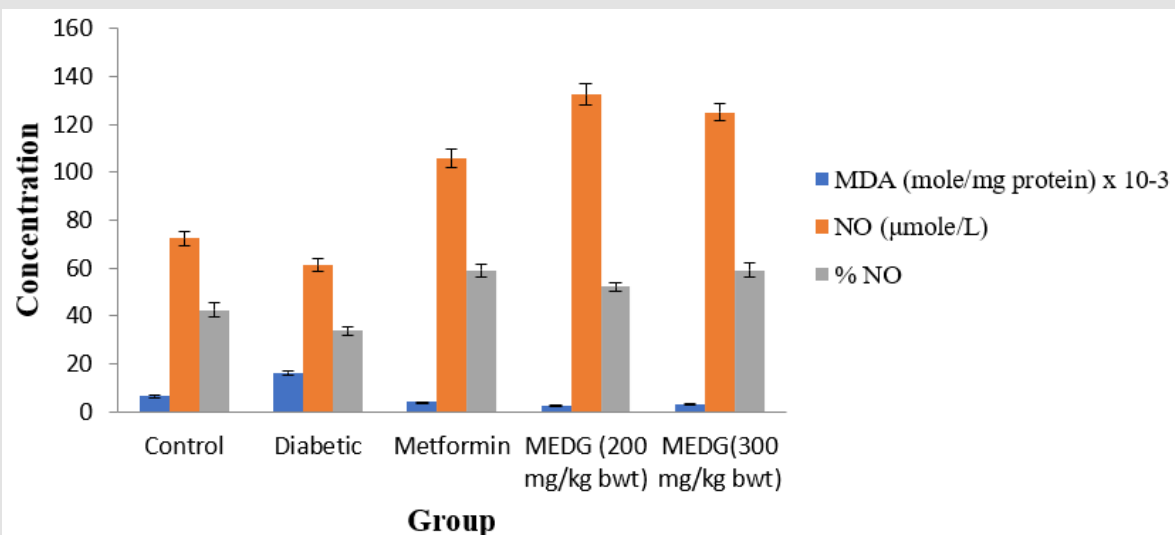
Note: Data are markers of oxidative stress and are expressed as mean ± SEM (n = 5).

Figure 2: Effect of MEDG Stem Bark on Oxidative Status in Rat Plasma.



Note: Data are activities of plasma α-amylase and glutathione reductase and are expressed as mean ± SEM (n = 5).

Figure 3: Effect of MEDG Stem Bark on Plasma Alpha-Amylase and Glutathione Reductase Activity.



Note: Data are plasma MDA and NO concentrations and are expressed as mean ± SEM (n = 5).

Figure 4: Effect of MEDG Stem Bark on Plasma MDA and NO Levels.

Discussion

Diabetes mellitus (DM) is a group of metabolic diseases characterized by chronic hyperglycemia that results from disturbed insulin secretion or function or both. Currently, many countries are on the verge of a global “diabetes epidemic”, which is rapidly spreading across the planet [1]. Chronic hyperglycemia in DM is accompanied by damage, dysfunction, and failure of various organs and tissues, development of micro- (retinopathy, nephropathy, and neuropathy) and

macrovascular (cardiovascular disorders) complications [24]. Reactive oxygen species (ROS) are chemically active oxygen-containing molecules generated in living systems. They are natural by-products of oxygen metabolism in all aerobic organisms. The main ROS types include superoxide, hydroperoxyl radical, singlet radical, hydroxyl radical, nitric oxide, peroxynitrite, amongst others [25]. Although ROS are primarily generated in mitochondria, but other alternative mechanisms contribute to their formation: NADPH-oxidase (NOX), immune reactions, xanthine oxidase, arachidonic acid metabolism,

amongst others [26]. These molecules are widely involved in processes of intracellular signaling and regulation of cell activity—apoptosis induction, adaptation to the effects of various factors, and immune response [27]. Moreover, ROS can stimulate inflammatory responses through protein kinases, transcription factors, and pro-inflammatory factors genomic expression [28]. Increased ROS accumulation leads to oxidative stress, which contributes to major cellular components (lipids, proteins, and DNA) damage.

The antioxidant defense system provides critical defense for the biological system by limiting the damaging effects of ROS. There are many antioxidant enzymes, including SOD, GPx, GR, catalase, paraoxanase (PON), amongst others [29]. In addition to enzymatic antioxidants, non-enzymatic antioxidant defense (ascorbate, tocopherols, retinol, carotenoids, reduced glutathione (GSH), melatonin, polyphenols, ceruloplasmin, carnosine, amongst others) also play crucial role in maintaining normal ROS levels. Under different pathological conditions, including DM, the redox balance can be disturbed that leads to negative consequences for the cell [2,30]. Many phytochemicals including phenolics, flavonoids, tannins, proanthocyanidins, and various plant extracts have been reported as antioxidants [31-33]. The aim of this study was to evaluate the effect of MEDG stem bark on oxidative stress in rat plasma caused by DM. The results obtained indicated that the activities of all the antioxidant enzymes and concentrations of GSH were significantly lower in diabetic group than in the control group, but they were increased by MEDG treatment. However, the concentrations of NO and MDA elevated by STZ were greatly reduced after treatment with the medicinal plant extract. These results are in agreement with reports of previous studies [34-45]. Studies have shown that plants rich in important bioactive compounds/phytochemicals are very useful medicinally [46-69].

Conclusion

The results obtained in this study have shown that MEDG stem bark can stimulate antioxidant defense system in rats challenged with STZ. In addition, the extract may serve as natural source of antioxidants.

Competing Interests

The authors declare that they have no conflict of interest.

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References

- (2012) Diabetes Report Card. Atlanta, GA: Centers for Disease Control and Prevention, U.S Department of Health and Human Services.
- Onoagbe I, Esekheigbe A (1999) Studies on the anti-diabetic properties of *Uvaria Chamae* in streptozotocin-induced diabetic rabbits. *Biokemistri* 9: 79-84.
- Rains JL, Jain SK (2011) Oxidative stress, insulin signaling, and diabetes. *Free Radic Biol Med* 50(5): 567-575.
- Tiedge M, Lortz S, Drinkgern J, Lenzen S (1997) Relation between antioxidant enzyme gene expression and antioxidative defense status of insulin-producing cells. *Diabetes* 46(11): 1733-1742.
- Robertson RP, Harmon J, Tran PO, Tanaka Y, Takahashi H, et al. (2003) Glucose toxicity in beta-cells: type 2 diabetes, good radicals gone bad, and the glutathione connection. *Diabetes* 52(3): 581-587.
- Brownlee M (2003) A radical explanation for glucose-induced beta cell dysfunction. *J Clin Invest* 112(12): 1788-1790.
- Rizzo MA, Piston DW (2003) Regulation of beta cell glucokinase by S-nitrosylation and association with nitric oxide synthase. *J Cell Biol* 161(2): 243-248.
- Tejedo J, Bernabe JC, Ramirez R, Sobrino F, Bedoya FJ, et al. (1999) NO induces a cGMP-independent release of cytochrome c from mitochondria which precedes caspase 3 activation in insulin producing RINm5F cells. *FEBS Lett* 459(2): 238-243.
- Abu OD, Imafidon KE, Obayuwana HO, Okuofu ED (2017) Phytochemical, proximate, and metal content analysis of *citrullus lanatus* (watermelon) seeds. *FUDMA Journal of Sciences* 2(2): 153-156.
- Dressler S, Schmidt M, Zizka G (2014) *Dialium guineense*: African Plants – A photo guide. Frankfurt/Main: Forschungsinstitut Senckenberg.
- Kar A (2007) Pharmacognosy and Pharmaco-biotechnology (Revised-Expanded (2nd Edn.)). New Age International Limited Publishers, New Delhi. pp. 332-600.
- Bero J, Ganfon H, Jonville MC, Frederich M, Gbaguidi F, et al. (2009) In vitro antiparasmodial activity of plants used in Benin in traditional medicine to treat malaria. *Journal of Ethnopharmacology* 122(3): 439-444.
- Abu OD, Onoagbe IO (2019) Biochemical effect of aqueous extract of *Dialium guineense* stem bark on oxidative status of normal Wistar rats. *International Journal of Clinical Biology and Biochemistry* 1(2): 15-18.
- Abu OD, Adeogun EF, Ebhohon SO (2019) Oral LD50 of total saponins and tannins isolated from *Dialium guineense* stem bark. *European Journal of Experimental Biology* 9(2): 11-13.
- Abu OD, Onoagbe IO (2021) Acute toxicity of aqueous and ethanol extracts of *Dialium guineense* stem bark. *Journal of Bioinnovation* 10(2): 427-432.
- Cohen G, Dembie CD, Marcus J (1970) Measurement of catalase activity in tissue extracts. *Analytic Biochemistry* 34: 30-38.
- Misra HR, Fridovich I (1972) The role of superoxide anions in the auto oxidation of epinephrine and a single assay for superoxide dismutase. *J Biol. Chem* 247: 3170-3175.
- Rotruck JT, Pope AL, Ganther HE, Swanson AB, Hafeman DG, et al. (1973) Selenium biochemical role as a component of glutathione peroxidase. *Science* 179(4073): 588-590.
- Abu OD, Ikponmwosa-Eweka O (2022) Evaluation of the Potential of Total saponins and Tannins of *Dialium guineense* Stem Bark in the Amelioration of Carbon Tetrachloride-Induced Renal Oxidative Stress. *SAU Science-Tech. Journal* 7(1): 42-50.
- Henry RJ, Sobel C, Beckman S (1957) Determination of serum protein by the Biuret reaction. *Anal Chem* 92(149): 1-5.
- Ellman GL (1959) Tissue sulphydryl groups. *Archive of Biochemistry and Biophysics* 82(1): 70-77.
- Guttridge JMC, Wilkins C (1982) Cancer dependent hydroxyl radical damage to ascorbic acid. Formation of thiobarbituric acid reactive product. *FEBS Lett* 137(2): 327-340.

23. Marcocci L, Packer L, Droy-Lefaix MT, Sekaki A, Gardes-Albert M (1994) Antioxidant action of Ginkgo biloba extract EGB 761. *Methods in Enzymology* 234: 462-475.
24. Harding JL, Pavkov ME, Magliano DJ, Shaw JE, Gregg EW (2019) Global trends in diabetes complications: a review of current evidence. *Diabetologia* 62(1): 3-16.
25. Sies H, Jones DP (2020) Reactive oxygen species (ROS) as pleiotropic physiological signalling agents. *Nat Rev Mol Cell Biol* 21(7): 363-383.
26. Forrester SJ, Kikuchi DS, Hernandes MS, Xu Q, Griendling KK, et al. (2018) Reactive oxygen species in metabolic and inflammatory signaling. *Circ Res* 122(6): 877-902.
27. Sies H (2020) Oxidative stress: concept and some practical aspects. *Antioxidants* 9(9): 852.
28. Mehta MM, Weinberg SE, Chandel NS (2017) Mitochondrial control of immunity: beyond ATP. *Nat Rev Immunol* 17(10): 608-620.
29. Ozougwu JC (2016) The role of reactive oxygen species and antioxidants in oxidative stress. *Int J Res Pharm Biosci* 3(6): 1-8.
30. Abu OD, Imafidon KE, Iribhogbe ME (2015) Biochemical effect of aqueous leaf extract of *Icacina trichanta* Oliv. on urea, creatinine and kidney oxidative status in CCl₄-induced Wistar rats. *Nigerian Journal of Life Sciences* 5(1): 85-89.
31. Abu OD, Okuo AV, Osemwota OF (2022) Extracts of *Dialium guineense* Stem Bark Ameliorates CCl₄-induced Oxidative Stress in Liver of Wistar Rats. *Biomedical Journal of Scientific and Technical Research* 46(2): 37297-7301.
32. Abu OD, Iyare HE, Ogboi KU (2022) Cardiac Oxidative Status in CCl₄-Exposed Rats Treated with Extracts of *Dialium guineense* Stem Bark. *Global Journal of Scientific Frontier Research* 22(1): 1-6.
33. Abu OD, Iyare HE, Ogboi KU (2022) Antioxidant Property of Total Saponins and Tannins of *Dialium guineense* Stem Bark in Rats Hearts Exposed to CCl₄. *Journal of Clinical Epidemiology and Toxicology* 3(3): 1-4.
34. Abu OD, Onoagbe IO, Obahiagbon O (2020) *In Vitro* Antioxidant Activities of Extracts of *Dialium Guineense* Stem Bark. *American Journal of Sciences and Engineering Research* 3(4): 68-75.
35. Abu OD, Onoagbe IO, Obahiagbon O (2020) Phenolic contents of extracts of *Dialium guineense* stem bark. *American Journal of Sciences and Engineering Research* 3(4): 92-96.
36. Abu OD, Onoagbe IO, Obahiagbon O (2020) Qualitative phytochemical screening and proximate analysis of *Dialium guineense* stem bark. *IAR Journal of Agriculture Research and Life Sciences* 1(4): 108-112.
37. Abu OD, Ezike TV, Ajuwa OI (2022) Cardioprotective property of extracts of *Dialium guineense* stem bark in rats exposed to CCl₄. *American Journal of Biomedical Science and Research* 16(6): 689-693.
38. Abu OD, Umar AB, Eiremiokhae CO (2022) Investigation of the Cardioprotective Capacity of aqueous Extract of *Icacina trichanta* Leaves in Rats Exposed to CCl₄. *Journal of Genetics and Cell Biology* 6(1): 322-328.
39. Abu OD, Onoagbe IO, Ekugum E (2022) Hepatotoxicity of Graded Doses of Ethanol Extract of *Dialium guineense* Stem Bark in Wistar Rats. *Journal of Pharmaceutical and Bio-Medical Sciences* 2(9): 347-352.
40. Abu OD, Onoagbe IO, Ohikhuare F (2022) Nephrotoxic Evaluation of Ethanol Stem Bark Extract of *Dialium guineense* in Normal Wistar Rats. *International Journal of Forensic Medicine* 4(2): 19-22.
41. Abu OD, Umar AB, Adekanle E (2022) Cardiotoxic Effect of Aqueous Extract of *Dialium guineense* Stem Bark in Wistar Rats. *East African Scholars Journal of Agriculture and Life Sciences* 5(9): 167-172.
42. Abu OD, Okuo AV, Ayele PE (2022) Pancreatotoxic Effect of Aqueous Extract of *Dialium guineense* Stem Bark in Wistar Rats. *International Journal of Novel Research in Life Sciences* 9(5): 31-37.
43. Patil MVK, Kandhare AD, Bhise SD (2012) Effect of aqueous extract of *Cucumis sativus* Linn. fruit in ulcerative colitis in laboratory animals. *Asian Pacific Journal of Tropical Biomedicine* 2(2): S962-S969.
44. Abu OD, Osime EC, Ngedaa OS (2023) Cardiac Oxidative Status in Diabetic Wistar Rats Exposed to Ethanol Extract of *Cucumis sativus*. *J Diagnostics and Case Reports* 4(2): 1-5.
45. Abu OD, Ojo I, Awhin EP (2023) Protective Property of Ethanol Extract of *C. sativus* on STZ-Induced Diabetic Rat Pancreas. *Biomedical Journal of Scientific and Technical Research* 52(2): 43613-43618.
46. Abu OD, Awhin EP, Ozedu ME (2023) Evaluation of Cardiovascular Disease Risk Factors in Diabetic Rats Administered Ethanol Extract of *Cucumis sativus* Fruit. *African Journal of Health, Safety and Environment* 4(1): 108-117.
47. Abu OD, Awhin EP, Iyare HE (2023) Assessment of Renal Function in Diabetic Wistar Rats Treated with Ethanol Extract of *Cucumis sativus*. *African Journal of Health, Safety and Environment* 4(1): 101-107.
48. Abu OD, Avenbuan SE, Osarhenomase EG (2023) Renal Oxidative Status in Diabetic Wistar Rats Administered Ethanol Extract of *Cucumis sativus* Whole Fruit. *Int J of Clinical Studies and Medical Case Reports* 30(1): 1-4.
49. Abu OD, Obaze GE, Egili S, Idehen IO (2023) Ethanol Extract of sativus Modulates the Activity of Glucose 6-phosphatase/ Aminotransferases and Levels of Lipids in Tissues of STZ- Induced Diabetic Rats. *Biomedical Journal of Scientific and Technical Research* 53(4): 44989-44994.
50. Abu OD, Osagie AO, Kolawole OM (2022) Ameliorative Effect of Extracts of *Dialium guineense* Stem Bark in CCL₄-Induced Kidney Dysfunction in Wistar Rats. *Biokemistri* 34(2): 34310-34316.
51. Abu OD, Iyare HE, Omoruyi IJ (2022) Toxic Responses of the Blood of Rats Exposed to Aqueous Extract of *Dialium guineense* Stem Bark. *FUDMA Journal of Science* 7(2): 117-120.
52. Abu OD, Imafidon KE, Obayuwana HO, Onodje S (2020) Quantitative phytochemical evaluation and phenolic contents of extracts of *Citrullus lanatus* seed. *Int J Bioorg Chem Mol Biol* 7: 31-35.
53. Abu OD, Onoagbe IO, Ojo I (2022) Dose response study of aqueous extract of *Dialium guineense* stem bark. *American Journal of Biomedical Science and Research* 15(2): 250-252.
54. Abu OD, Onoagbe IO, Ojo I (2022) Dose response of total saponins isolated from the stem bark of *Dialium guineense*. *Journal of Advances in Plant Biology* 1(4): 1-6.
55. Abu OD, Onoagbe IO, Ojo I (2021) Determination of effective dose for ethanol extract of *Dialium guineense* stem bark. *Journal of Medical Research and Case* 3(2): 1-4.
56. Abu OD, Ikponmwosa Eweka O (2022) Potential of Extracts of *Dialium guineense* Stem Bark in the Mitigation of Carbon Tetrachloride-induced Renal Oxidative Stress. *BIU Journal of Basic and Applied Sciences* 7(1): 62-69.
57. Abu OD, Ikponmwosa Eweka O (2022) Potential of Total Saponins and Tannins Isolated from the stem bark of *Dialium guineense* in the Amelioration of Kidney Dysfunction Caused by CCl₄. *Journal of Basic and Applied Medical Sciences* 2(1): 6.
58. Abu OD, Onoagbe IO, Ojo I (2021) Graded and quantal dose response of total tannins isolated from the stem bark of *Dialium guineense*. *Advanced Research Journal of Medicine and Clinical Science* 8(10): 699-703.

59. Abu OD, Ojo I, Ezike TV (2023) Methanol Fraction of Ethanol Extract of *Dialium guineense* Stem Bark Mitigates STZ-Induced Oxidative Stress in Rat Liver. Biomedical Journal of Scientific and Technical Research 51(2): 42594-42600.
60. Abu OD, Awhin EP, Ohikhuare F (2023) Effect of Methanol Fraction of Ethanol Extract of *Dialium guineense* Stem Bark on Cardiovascular Disease Risk Factors in Diabetic Rats. Journal of Biology and Medicine 4(1): 128.
61. Abu OD, Okuo AV, Egili S, Idehen IO, Esedebe M1, et al. (2023) Methanol Fraction of Ethanol Extract of *Dialium guineense* Stem Bark May Alter the Activity of Glucose 6- phosphatase/Aminotransferases and Levels of Lipids in Tissues of Diabetic Wistar Rats. International Journal of Research and Scientific Innovation 10(12): 523-532.
62. Abu OD, Alegun O, Ifekwe JC (2023) Renal Oxidative Status in Diabetic Wistar Rats Administered Methanol Fraction of Ethanol Extract of *Dialium guineense*. Medical and Clinical Case Reports Journal 1(1): 1-13.
63. Abu OD, Ojo I, Awhin EP, Iyorah I (2024) Methanol Fraction of Ethanol Extract of *Dialium guineense* Stem Bark Reduces Oxidative Stress in STZ-Induced Diabetic Rat Pancreas. International Journal of Forensic Medicine 6(2): 12-19.
64. Abu OD, Awhin EP, Ohikhuare F, Osamudiamen EE (2024) Investigation of the biochemical effect of aqueous extract of *Dialium guineense* stem bark on haematological parameters in rats. Biomedical Journal of Scientific and Technical Research 58(1): 49990-49997.
65. Abu OD, Umar AB, Ekperusi SE, Ohikhuare F (2024) Assessment of Cardiac UOxidative Status of Diabetic Wistar Rats Exposed to Methanol Fraction of Ethanol Extract of *Dialium guineense* Stem Bark. Biomedical Journal of Scientific and Technical Research 57(2): 49002-49009.
66. Abu OD, Awhin EP, Iyoha AI, Chukwuma AU (2024) Hepatotoxicity of Aqueous Extract of *Dialium guineense* Stem Bark. Biomedical Journal of Scientific and Technical Research 55(2): 46902-46907.
67. Abu OD, Okolie NP, Ekperusi SE, Marcel LI (2024) Acute Toxicity Study of Ethanol Extract of *Phyllanthus amarus* Leaves in Wistar Albino Rats. Biomedical Journal of Scientific and Technical Research 57(2): 49010-49013.
68. Abu OD, Okuo AV, Egili S, Idehen IO, Eze Nwaobasi OP, et al. (2024) Plasma oxidative status of diabetic rats treated with ethanol extract of *C. sativus* fruit. International Journal of Biology and Medicine Open Access 1(1): 1-7.
69. Iyoha AI, Onoagbe IO, Abu OD (2023) Effects of aqueous and methanolic leaf extracts of *Lonchocarpus cyanencens* leaf on oxidative status in normal albino Wistar rats. Nigerian Journal of Life Sciences 13(1,2): 7-10.

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