

# The Biology of the Facultative Halophyte *Atriplex patula* L.

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## ABSTRACT

*Atriplex patula* L. is a facultative halophyte that tolerates low to moderate levels of salinity and occurs on the margins of coastal salt marshes, roadsides, in agricultural fields, and in other areas where disturbance has occurred. The genus *Atriplex* L. is represented by about 250 species globally and has recently been included in the family Amaranthaceae subfamily Chenopodioidae. *Atriplex patula* was possibly introduced in North America from Europe, Asia or North Africa in the 18th century. Because the stems and leaves accumulate and store heavy metal pollutants, *A. patula* has been suggested for use in habitat remediation.

**Keywords:** Spear Orach; Taxonomy; Geographical Distribution; Dimorphic Seeds; Germination; Facultative Halophyte; Physiological Ecology; Range of Habitats

## Introduction

The genus *Atriplex* L. is represented by about 250 species globally. Molecular and morphological data have provided some evidence supporting the inclusion of the Chenopodiaceae in the family Amaranthaceae with the former family Chenopodiaceae now listed as a subfamily Chenopodioidae [1]. However, Welsh et al. [2] and others still use the *nomen* Chenopodiaceae in systematic treatments. *Atriplex patula* L. (spear orach, spearscale) is a facultative halophyte that tolerates low to moderate levels of soil salinity [3]. It is a widespread weedy species that occurs in agricultural fields, on roadsides, on the margins of coastal salt marshes, and in other areas where disturbance has occurred. *Atriplex patula* produces dimorphic seeds which allows the species to be established in unstable habitats [4, 3]. Welsh [1] indicated that *A. patula* was possibly introduced in North America from Europe, Asia, or North Africa in the 18<sup>th</sup> century, and its occurrence has been documented in at least 10 provinces in Canada and 29 states in the U.S.A. Chetina et al. [5] have reported that *A. patula* occurs in saline sites at 59° N near Perm Krai, Russia, and De Cauwer et al. [6] noted that this species is a weed control problem in sugar beet fields in Belgium. *Atriplex patula* has been suggested for use in

habitat remediation because stems and leaves accumulate and store heavy metal pollutants [7]. Herein, we summarize important aspects of this widespread facultative halophyte.

## Synonyms, Taxonomy, Variation

Synonymy of the *Atriplex patula* complex is in a state of confusion. Synonyms of *Atriplex patula* L. (spear orach, spreading orache, spear-scale) in North America have primarily been listed as *Atriplex hastata* L. subsp. *patula* (L.) S. Pons, *A. hastata* var. *patula* (L.) Farwell, and *Teutiopsis patula* (L.) Selak [1], whereas Correll and Correll [8] listed *A. patula* L. as *A. patula* L. var. *hastata* (L.) Gray. The nomens *A. prostrata* Boucher and *A. hastata* L. *sensu* Aellen *non* L. have also been listed as synonyms by Stalter and Lamont [9]. The following taxonomic description of *A. patula* is summarized from treatments of the species by Correll and Correll, [8], Stalter, [10], Taylor and Wilken, [11], and Welsh, [1]. *Atriplex patula* is a monoecious annual with a root system that typically extends between 10 to 18 cm below the substrate surface. Stems are erect and branched, glabrous to scaly, and range from 30 cm to 1.5 m in height. Leaves are opposite below and alternate above.

Petioles of lower leaves are up to half as long as the blades. Blades are bright green, 2.5 to 7.0 cm long and nearly as broad, truncate at the base, and acute or obtuse at the apex. The staminate flowers bearing 5 tepals and 5 stamens are borne in a terminal spike or panicle. The pistillate flowers are usually borne in few-flowered axillary or spike-like cymes. Pistillate flowers have 2 stigmas, and the base of the ovary is enclosed in a pair of bracts often reddish at maturity and up to 7 mm long. Seeds are dimorphic. Brown seeds are 2.5 to 4.0 mm wide, and black seeds are 1.0 to 2.0 mm wide. Chromosome numbers for the *A. patula* complex are  $2n = 18$  and  $2n = 36$ . In the Netherlands, Van Der Meyden [12] tentatively recognized *A. patula* L. and *A. hastata* L. as separate species. However, he noted that there are no sharp distinctions between the two taxa, and numerous overlapping morphological variations exist in leaf blade morphology between the two entities. Van Der Meyden [12] stated that different ploidy levels occur in the complex. *Atriplex hastata* is a diploid and *A. patula* is a tetraploid.

## Geographic Distribution

*Atriplex patula* is an annual, weedy, facultative halophyte that has a wide distributional range in North America, Europe, North Africa, and Asia. Correll and Correll [8] reported its range in North America from Newfoundland westward to British Columbia in Canada, and from Newfoundland to South Carolina in the U.S.A. Stalter et al. [13] reported the occurrence of the species in coastal sites in northern Florida. On the Pacific coast of North America, it occurs from British Columbia to California. Ungar [3] indicated that it occurs in low to moderately saline sites in Kansas and Nebraska, and Correll and Correll [8] and Proffitt et al. [14] stated that *A. patula* is often present in saline sites in Texas and Louisiana. *Atriplex patula* is widespread in Europe. De Cauwer et al. [6] reported that *A. patula* is a weed control problem in sugar beet fields in Belgium, and Chetina et al. [5] reported the occurrence of this species in salinized alluvial soils at 59° N latitude at Perm Krai, Russia.

## Range of Habitats

*Atriplex patula* is a weedy, facultative halophyte that occurs in a wide variety of soil types including sandy loams and clays [6], salt impacted alluvial soils [5, 15], and water-logged soils [16]. In sugar beet fields in Belgium, De Cauwer et al. [6] reported that the species is less common in acid sandy soils and is less tolerant to drought than to water-logging. Ungar [3] indicated that *A. patula* is a summer annual in the Central U.S.A. and can tolerate moderately saline soils under field conditions ranging from 2 ppt to 13 ppt total salts. In North Carolina, Stalter and Lamont [9] stated that habitats for *A. patula* are beaches and upper margins of brackish and salt marshes. In New England salt marshes, Rand [17-19] reported that this species is usually found in bare areas in the mid to high marsh in topographic zones usually dominated by *Juncus gerardii* or *Iva frutescens*. She noted that the annual halophyte *Salicornia europaea* may compete with *A. patula* in this harsh physical environment. Proffitt et al. [14] found a simi-

lar case in salt marshes in Louisiana where *A. patula* occurs in bare patches within sites dominated by *Sporobolus alterniflorus* (*Spartina alterniflora*).

## Physiological Ecology

*Atriplex patula* has Kranz anatomy in leaves which is indicative of the associated  $C_4$  carbon fixation pathway of photosynthesis [1]. The  $C_4$  pathway increases efficiency of  $CO_2$  usage and subsequent carbon fixation. Salinity is an important factor that affects imbibition of water in seeds, germination, root elongation, and overall plant vitality [20]. NaCl in the substrate affects osmotic balance and specific ions. Salinization influences the production of the low molecular weight amino acid proline in *A. patula* leaves. Proline acts as an antioxidant and protects cellular protein structure and ultimately increases *A. patula* resistance to salt in acidic soil conditions [5].

## Dimorphic Seeds and Germination

Two types of seeds occur in *A. patula*. One is black and ranges in size from 1.0 to 2.0 mm in diameter, and the other seed type is brown and ranges in diameter from 1.3 to 3.0 mm [4, 3, 1]. Large brown seeds have three times the mass of smaller black seeds, a thin seed coat, a larger embryo, and absorbs water 100 times faster than unscarified black seeds [20-21]. Nurse et al. [21] stated that seed dimorphism enhances survival of the species in variable habitats. Larger brown seeds have a limited dormancy period and germinate rapidly in spring. However, brown seeds are more susceptible to fungal attack than black seeds [21]. Ungar [22] reported that black seeds have a hard seed coat, a long dormancy period, and scarification is required to break dormancy. In the laboratory, unscarified black seeds did not germinate. However, 73% of scarified black seeds germinated [4, 22]. Black seeds have greater resistance to high salinity stress than brown seeds and retain their viability longer than brown seeds [21]. In the laboratory, Ungar [22] found that seeds germinated and seedlings survived salinity concentrations up to 20 ppt. Germination was reduced at salinity values greater than 10 ppt, and germination declined to 17% of controls at 20 ppt. Ungar [22] also noted that seeds are less inhibited to high salinity levels that growing seedlings. In Oregon, Keammerer and Hacker [23] stated that *A. patula* seed viability was 91% in the high intertidal zone, but they did not mention seed dimorphism.

## Competition and Herbivores

*Atriplex patula* does not fare well in competition with taller or more aggressive species in salt marshes. Minchinton et al. [24] reported that the common reed (*Phragmites australis*), in the upper margins of New England salt marshes, modifies soil and light conditions and produces a heavy cover of litter. These factors exclude the perennial *Solidago sempervirens* and the annual *A. patula* from this topographic zone. *Atriplex patula* seedling establishment and survival of mature plants only occurs at sites that have not been invaded by

*P. australis*. A chrysomelid beetle (*Erynephala maritima*) is a herbivore that damages *A. patula* and the succulent annual halophyte *Salicornia europaea* in salt marshes in Rhode Island [25]. Damage to *A. patula* is extensive at mid-tidal elevations which are frequently inundated [17]. She also found that *A. patula* plants are more extensively damaged when they are not present in dense stands of *S. europaea* [17]. In field plots cleared of *S. europaea* beetle herbivory reduced *A. patula* survival by about 50% and reduced reproduction by 37% [18, 19].

## Phytoremediation

*Atriplex patula* is a species that removes significant amounts of Na ions from salt-impacted soils and stores salt in cellular vacuoles in stems and leaves [15]. They noted that the deep root system improves soil quality and soil permeability in oil field brine spill sites in Saskatchewan, Canada. Mature plants are removed of spill sites and are disposed off-site. High seed production insures a standing crop in the next growing season.

Vickerman et al. [7] examined *A. patula* for potential habitat improvement and phytoremediation of the heavy metal Se. They found that phytoremediation was greatest in saline sites having low to moderate levels of sulfate. They also indicated that mature plants could be mechanically harvested and disposed off-site.

## Management and Role as an Allergen

*Atriplex patula* is an aggressive annual weed in sugar beet fields in Belgium. Various herbicides have been used to control the species. However, this species exhibits high intraspecific variation in herbicide sensitivity and weed control by herbicides has been shown to be difficult [6]. *Atriplex* species produce extensive amounts of wind-blown pollen and may play a role in allergic sensitization [26]. Ferrer et al. [27] noted that pollen of *A. patula* extracts should be used in a battery of potential allergens for use in diagnosis and treatment of respiratory allergies in patients.

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## References

- Welsh SL (2003) *Atriplex* L. In: Flora of North America Editorial Committee (Edt.), Magnoliophyta Caryophyllidae (Part 1): Chenopodiaceae. Flora of North America North of Mexico. Oxford: Oxford University Press pp. 322-324.
- Welsh SL, CW Crompton, E Clemants (2003) Chenopodiaceae Ventenat. In: Flora of North America Editorial Committee (Edt.), Magnoliophyta Caryophyllidae (Part 1): Chenopodiaceae. Flora of North America North of Mexico. Oxford: Oxford University Press 4: 258.
- Ungar IA (1996) Effect of salinity on seed germination, growth, and ion accumulation of *Atriplex patula* (Chenopodiaceae). American Journal of Botany 83(5): 604-607.
- Drysdale F R (1973) Variation of seed size in *Atriplex patula* var. *hastata* (L.) Gray. Rhodora 75(801): 106-110.
- Chetina OA, Eremchenko OZ, Pakhorukov IV (2023) Accumulation of low-molecular-weight compounds in plants in response to technogenic soil salinization. Russian Journal of Ecology 54(2): 88-96.
- De Cauwer B, Cardinael A, Claerhout S, Manderyck B, Reheul D (2018) Differential sensitivity of *Atriplex patula* and *Chenopodium album* to sugar beet herbicides: A possible cause for the upsurge of *A. patula* in sugar beet fields. Weed Research 58(2): 99-111.
- Vickerman DB, Shannon MC, Bañuelos GS, Grieve CM, Trumble JT, et al. (2002). Evaluation of *Atriplex* lines for selenium accumulation, salt tolerance, and suitability for a key insect agricultural pest. Environmental Pollution 120(2): 463-473.
- Correll DS, Correll HB (1972) Aquatic and Wetland Plants of the Southwestern United States. Washington, D.C.: Environmental Protection Agency, pp. 1775.
- Stalter R, Lamont EE (1999) Vascular flora of Cape Lookout National Seashore and Bogue Banks, North Carolina. Journal of the Elisha Mitchell Society 115(4): 213-235.
- Stalter R (1974) Vegetation of coastal dunes of South Carolina. Castanea 39(1): 95-103.
- Taylor D, Wilken DH (1993) *Atriplex*. In: The Jepson Manual: Higher Plants of California. In: Hickman (Edt.), Berkeley, California: University of California Press pp. 501-505.
- Van Der Meyden R (1970) Biosystematic notes on *Atriplex patula* L. *A. hastata* L. and *A. littoralis* L. (Chenopodiaceae). Blumea 18(1): 53-63.
- Stalter R, Leyva M, Kincaid DT (1999) The flora of Indian Shell Rings from coastal South Carolina to northern Florida. Sida 18(3): 298-875.
- Proffitt CE, Chiasson RL, Owens AB, Edwards KR, Travis SE (2005) *Spartina alterniflora* genotype influences facilitation and suppression of high marsh species colonizing an early successional salt marsh. Journal of Ecology 93(2): 404-416.
- Young MA, Rancier DG, Roy JL, Lunn SR, Armstrong SA, et al. (2011) Technical note: Seedling condition of the halophyte *Atriplex patula* for optimal growth on a salt impacted site. International Journal of Phytoremediation 13(7): 674-680.
- Magnanti M, Weaver S, Downs M (2005) Responses of spreading orach (*Atriplex patula*) and common lamb quarters (*Chenopodium album*) to soil compaction, drought, and water logging. Weed Science 53(1): 90-96.
- Rand TA (2002) Variation in insect herbivory across a salt marsh tidal gradient influences plant survival and distribution. Oecologia 132(4): 549-558.
- Rand TA (2003) Herbivore-mediated apparent competition between two salt marsh forbs. Ecology 84(6): 1517-1526.
- Rand TA (2004) Competition, facilitation, and compensation for insect herbivory in an annual salt marsh forb. Ecology 85(7): 2046-2052.
- Katembé WJ, Ungar IA, Mitchell JP (1998) Effect of salinity on germination and seedling growth of two *Atriplex* species (Chenopodiaceae). Annals of Botany 82(2): 167-175.
- Nurse RE, Reynolds WD, Doucet C, Weaver SE (2008) Germination characteristics of the dimorphic seeds of spreading orache (*Atriplex patula*). Weed Science 56(2): 216-223.
- Ungar IA (1971) *Atriplex patula* var. *hastata* seed dimorphism. Rhodora 73(796): 548-551.

23. Keammerer HG, Hacker SD (2013) Negative and neutral marsh plant interactions dominate in early life states and across physical gradients in an Oregon estuary. *Plant Ecology* 214(2): 303-315.
24. Minchinton TE, Simpson JC, Bertness MD (2006) Mechanisms of exclusion of native coastal marsh plants by an invasive grass. *Journal of Ecology* 94(2): 342-354.
25. Rand TA (1999) Effects of environmental content on susceptibility of *Atriplex patula* to attack by herbivorous beetles. *Oecologia* 121(1): 39-46.
26. Fuller TC, Mc Clintock EM (1986) *Poisonous Plants of California*. Berkley, California: University of California Press pp. 433.
27. Ferrer L, Carnés J, Rojas-Hijazo B, Lopez-Matas MA, Sobrevía MT, et al. (2010) Assessing degree of flowering implicates multiple Chenopodiaceae/Amaranthaceae species in allergy. *International Archives of Allergy and Immunology* 158(1): 54-62.

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